

A Convenient Synthesis of Oligonucleotides with a 3'-Phosphoglycolate and 3'-Phosphoglycaldehyde Terminus

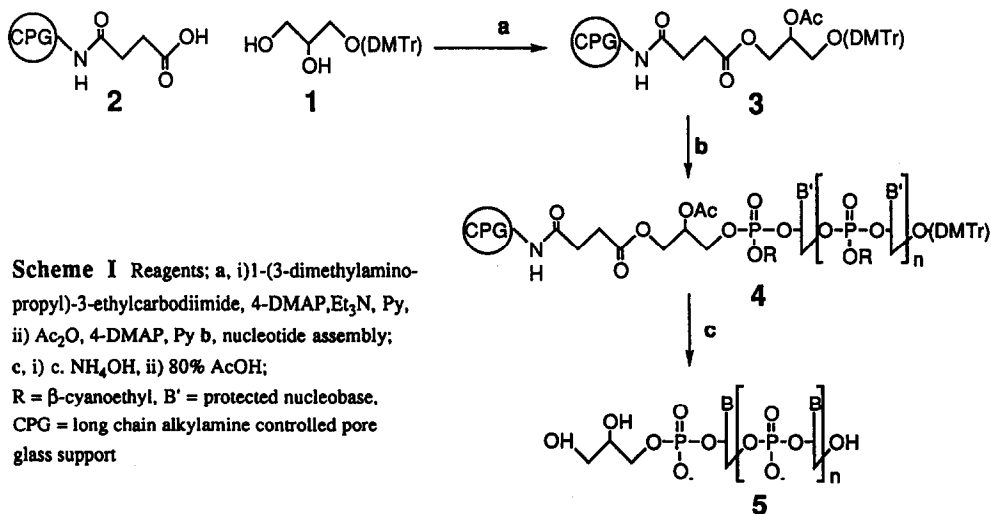
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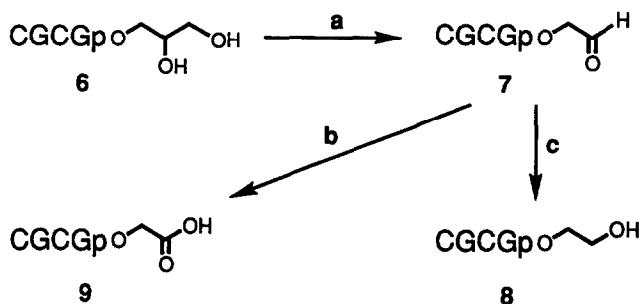
Abstract: A simple and rapid method for solid phase synthesis of oligonucleotides carrying a 3'-phosphoglycerol terminus was developed. The modified oligonucleotide was readily oxidized by NaIO_4 to a 3'-phosphoglycaldehyde. The treatment of this aldehyde with NaClO_2 afforded a 3'-phosphoglycolate.

Chemically synthesized oligonucleotides have been widely used in molecular biology and physicochemical studies of nucleic acids due to the recent rapid advance in the solid phase synthesis of oligonucleotides¹. Synthetic oligonucleotides that can be obtained with defined sequences are also useful for DNA-drug interaction study. Some DNA-targeting molecules such as bleomycin², the enediyne antibiotics^{3,4} and transition metal complexes⁵ cleave DNA strands in a radical process that includes hydrogen abstraction from the deoxyribose moieties of DNA. Identification of these DNA fragments produced by the drugs is



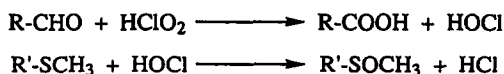
important and useful for the characterization not only of sequence specificity but of regiospecificity for hydrogen abstraction from the deoxyribose moiety as well⁶. Now we report a convenient method for the synthesis of oligonucleotides carrying 3'-phosphoglycolate³ and 3'-phosphoglycaldehyde^{6,7} terminus formed by abstraction of 4'- and 3'-hydrogen, respectively on deoxyribose moieties.

Mono-(dimethoxytrityl) glycerol(1) readily obtained from glycerol by treatment with dimethoxytrityl chloride/pyridine was condensed to the succinylated long chain alkylamine-controlled pore glass support (succinylated LCAA-CPG, 2), synthesized according to the literature procedure⁸, in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide. The resulting resin was acetylated with acetic anhydride/4-dimethylaminopyridine/pyridine to protect the 2-hydroxy group of the glycerol moieties and unreacted amino groups on the LCAA-CPG resin. The unreacted carboxyl groups on the resin were capped according to the literature procedure⁸. The common coupling cycle was repeated on the glycerol resin (3) by the β -cyanoethylphosphoramidite method until the desired oligonucleotide carrying a 3'-phosphoglycerol terminus (4) was obtained. Following conventional deblocking and purification, the chromatographically homogeneous tetranucleotide, d(CGCG) modified at a 3'-terminus (6) was quantitatively oxidized with NaIO₄ (final concentration; 5 mM)⁹ for 1.5 h at 0°C to give a 3'-phosphoglycaldehyde (7) as shown in Figure 1A and 1B. After the addition of L-methionine to the mixture to quench excess of NaIO₄¹⁰, the product was readily reduced to the corresponding alcohol (8) with NaBH₄ (Figure 1C). This confirms the structure of the aldehyde. Subsequent oxidation of the aldehyde with a NaClO₂ system¹¹ afforded 3'-phosphoglycolate (9)¹²



Scheme II Reagents; a, NaIO₄; b, NaClO₂; c, NaBH₄

as shown in Figure 1D and 1E. L-Methionine and its sulfoxide produced by quenching NaIO₄ could also serve as scavengers of HOCl generated by this oxidation reaction.



The tetranucleotide, d(CGCG) having the 3'-phosphoglycolate terminus (9) synthesized in this way was cochromatographed with a major bleomycin-mediated degradation product¹³ of d(CGCGCG) produced *via* H4' abstraction at the C(5) residue (Figure 1F).

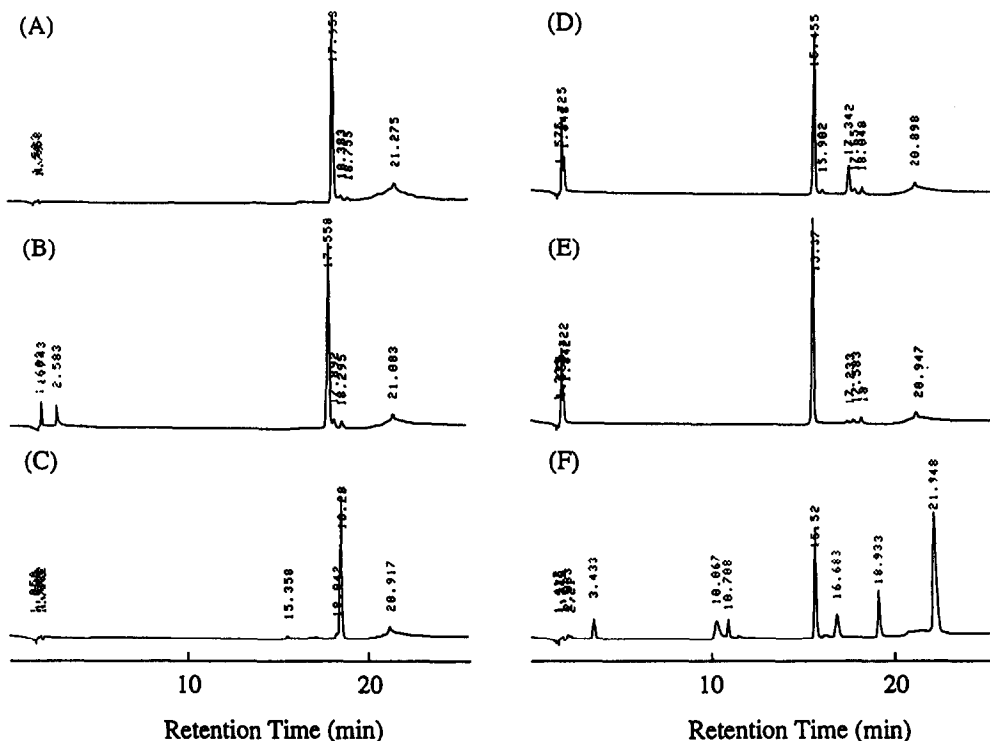


Figure 1. HPLC analysis of **6** and its conversion to **7**, **8**, and **9**, and that of bleomycin-mediated degradation products of d(CGCGCG). 3'-Phosphoglycerol tetranucleotide **6** (A) was converted to 3'-phosphoglycaldehyde **7** by treating with NaIO₄ at 0°C for 1.5 h (B). The treatment of **7** with NaBH₄ afforded the corresponding alcohol **8** (C). The aldehyde **7** was oxidized with NaClO₂ at 25°C for 2 h (D) and 5 h (E). The resulting 3'-phosphoglycolate was comigrated with the major product of the bleomycin-mediated strand cleavage reaction¹³ of d(CGCGCG)(F). The peak at 21.9 min in Fig. 1F corresponds to unreacted d(CGCGCG). Analysis was carried out on a μ Bondasphere C18 100Å column (3.9 x 150 mm); elution was performed with a linear gradient of acetonitrile 0-10% for 20 min containing 50 mM ammonium formate at a flow rate of 0.8 mL/min with 260 nm detection.

This method thus makes possible the direct identification of DNA degradation products produced by DNA damaging agents *via* H4' and H3' abstraction. Oligonucleotides with the 3'-phosphoglycaldehyde terminus should be useful for immobilization and attachment of oligonucleotides to polymer supports, proteins and fluorescent dyes *via* the glycaldehyde terminus.

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References and Notes

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10. The aldehyde was stable in this solution at least for a week at 4°C.
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12. Typically, an aqueous solution of a 3'-phosphoglycerol oligonucleotide was treated with 0.2 M NaIO₄ (5 mM final concentration) and the solution was kept at 0°C for 1.5 h. After the addition of 0.2 M L-methionine (7.5 mM final concentration), the mixture was kept at 0°C for additional 30 min. To this solution were added potassium phosphate buffer pH 4.0 (20 mM final concentration) and NaClO₂ (400 μM final concentration). The mixture was kept at 25°C. The reaction was complete within 5 h, and excess of NaClO₂ was quenched by Na₂SO₃.
13. The reaction mixture contained 200 μM d(CGCGCG), 200 μM bleomycin and 50 mM sodium phosphate pH 7.0. The reaction was initiated by the addition of Fe(II)(NH₄)₂(SO₄)₂ (final concentration 200 μM), incubated at 0°C for 30 min and then analyzed by HPLC.

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